# **Regular Article**

# Histogram Analysis of Pharmacokinetic Parameters by Bootstrap Resampling from One-point Sampling Data in Animal Experiments

Seiji Takemoto, Kiyoshi Yamaoka\*, Makiya Nishikawa and Yoshinobu Takakura

Department of Biopharmaceutics and Drug Metabolism, Graduate School of Pharmaceutical Science, Kyoto University, Kyoto, Japan

Full text of this paper is available at http://www.jstage.jst.go.jp/browse/dmpk

**Summary:** A bootstrap method is proposed for assessing statistical histograms of pharmacokinetic parameters (AUC, MRT, CL and  $V_{ss}$ ) from one-point sampling data in animal experiments. A computer program, MOMENT(BS), written in Visual Basic on Microsoft Excel, was developed for the bootstrap calculation and the construction of histograms. MOMENT(BS) was applied to one-point sampling data of the blood concentration of three physiologically active proteins (<sup>111</sup>In labeled Hsp70, Suc<sub>20</sub>-BSA and Suc<sub>40</sub>-BSA) administered in different doses to mice. The histograms of AUC, MRT, CL and  $V_{ss}$  were close to a normal (Gaussian) distribution with the bootstrap resampling number (200), or more, considering the skewness and kurtosis of the histograms. A good agreement of means and SD was obtained between the bootstrap and Bailer's approaches. The hypothesis test based on the normal distribution clearly demonstrated that the disposition of <sup>111</sup>In-Hsp70 and Suc<sub>20</sub>-BSA was almost independent of dose, whereas that of <sup>111</sup>In-Suc<sub>40</sub>-BSA was definitely dose-dependent. In conclusion, the bootstrap method was found to be an efficient method for assessing the histogram of pharmacokinetic parameters of blood or tissue disposition data by one-point sampling.

# Key words: bootstrap method; pharmacokinetics; histogram analysis; hypothesis testing; sparse sampling; Monte Carlo

#### Introduction

A certain model is assumed in pharmacokinetic studies (PK) which describe the relationship of drug absorption, distribution and elimination to a drug concentration after administration. An attempt is made in PK to evaluate the relationship between the disposition characteristics of a drug and its parameters calculated, based on a particular model. Much interest in PK involves comparing the parameters between two groups of animals under different experimental conditions. In order to identify any statistically significant differences, one time course must be obtained from one animal in the common method.

However, apart from blood, the repeated sampling of tissues from an individual experimental animal is quite difficult, and groups of animals in small numbers are euthanized at each sampling time. This conventional "one-point sampling" method has been widely used in various experimental settings in preclinical studies. In such experiments, pharmacokinetic parameters are calculated using the average values of concentrations in the blood or in organs of three or four experimental animals. Since the variances of the parameters are not assessed in this conventional analysis, it is impossible to statistically compare the pharmacokinetic parameters in two animal groups. When the statistical histogram of AUC is approximated by the normal (Gaussian) distribution, Bailer's approach can be adopted as a "gold standard" to estimate the standard error of AUC. The linearity and the normality of AUC distribution are implicitly assumed in Bailer's approach.<sup>1)</sup> If the parameters are given by a nonlinear function of concentrations, or the statistical distributions of parameters

Received; February 2, 2006, Accepted; August 23, 2006

<sup>\*</sup>To whom correspondence should be addressed: Kiyoshi YAMAOKA, Department of Biopharmaceutics and Drug Metabolism, Graduate School of Pharmaceutical Science, Kyoto University, 46-29 Shimoadachi-cho, Yoshida, Sakyo-ku, Kyoto 606-8501, Japan. Tel. +81-75-753-4615, Fax. +81-75-753-4614, E-mail: yamaoka@pharm.kyoto-u.ac.jp

Abbreviations used are: Hsp70, heat shock protein 70; Suc-BSA, succinylated bovine serum albumin; AUC, the area under the concentration curve; MRT, mean residence time; CL, total clearance;  $V_{ss}$ , volume at steady-state

are unknown, Bailer's method has a limitation.

In the paper (1979) entitled "BOOTSTRAP **METHODS:** ANOTHER LOOK AT THE JACKKNIFE", Efron proposed a 'bootstrap method', a statistical interval analysis, by means of Monte Carlo simulation.<sup>2,3)</sup> The bootstrap method has been widely adopted in many areas with the rapid development of computer power and its simple algorithm compared with that of the jackknife. The basis of the bootstrap method involves resampling from the observed data and construction of histogram on computer. The bootstrap method has been applied to estimating the confidence intervals for population pharmacokinetic parameters by resampling n time courses from n population time courses permitting replacement.<sup>4-8)</sup> The bootstrap method was also proposed for assessing the bioequivalence between two drug formulations in a draft guidance from the United States Food and Drug Administration (FDA).<sup>9-12)</sup> This approach was also implemented in a computer program for linkage analysis, MENDEL4, which was developed for analyzing genetically discrete traits in pedigree and population data sets.<sup>13)</sup>

The evaluation method presented here is based on the bootstrap approach for assessing the histograms of pharmacokinetic parameters for data collected by a onepoint sampling method in animal experiments. We have developed a computer program, MOMENT(BS), that provides the histograms of the parameters by the bootstrap method, including area under the concentration curve (AUC), mean residence time (MRT), total clearance (CL), and volume of distribution at steady-state (V<sub>ss</sub>). MOMENT(BS) provides the mean, standard deviation (SD), skewness (SK) and kurtosis (KT) of the parameters by means of bootstrap resampling, in addition to the mean and SD and standard error (SE) by expanding Bailer's method. We selected, as model drugs, three physiologically active proteins: mouse recombinant heat shock protein 70 (Hsp70)<sup>14-18)</sup> and two succinylated bovine serum albumins (Suc<sub>20</sub>-BSA and Suc<sub>40</sub>-BSA).<sup>19,20)</sup> This aim of this study was to carry out a statistical comparison of the pharmacokinetic parameters between two animal groups under different conditions by means of a hypothesis test.

#### **Materials and Methods**

#### Proteins and tissue distribution experiments

Mouse recombinant heat shock protein 70 (Hsp70), succinylated bovine serum albumin (the number of succinic anhydride units conjugated to BSA was 20 or 40; Suc<sub>20</sub>-BSA and Suc<sub>40</sub>-BSA) were radiolabeled with <sup>111</sup>In using the bifunctional chelating agent DTPA anhydrate. <sup>111</sup>In-Hsp70, Suc<sub>20</sub>-BSA and Suc<sub>40</sub>-BSA were injected into the tail vein of mice at different doses. The radioactivity in plasma was measured at the indicated times.<sup>21-25)</sup>

## Data analysis

**Bootstrap algorithm:** The procedures based on the bootstrap method for one-point sampling are as follows.

Step1. Construction of time course of blood level or tissue concentrations, which consist of three or four points at each time, from data obtained by one-point sampling in animal experiments (one point at each time is collected from one animal).

Step2. Selection of one point from three or four points at each time, permitting replacement using random number, and construction of pseudo-profile.<sup>26)</sup>

Step3. Calculation of pharmacokinetic parameters (statistics;  $\phi$ ) from time course obtained in Step2.

Step4. Construction of the histograms and calculation of moment characteristics (mean, SD, SK and KT) of  $\Phi$  with resampling of bootstrap number (B) times.

Step5. Assessment of histograms, including normal distribution and log-normal distribution.

Step6. Comparison of  $\Phi$  between two animal groups depending on type of statistical distribution.

To perform the above procedures, a computer program, MOMENT(BS), was developed, programmed in Visual Basic on Microsoft Excel. When concentrations in an organ are measured, AUC and MRT can be replaced by  $AUC_i$  and  $MRT_i$  in organ i, respectively.

The means  $(\bar{\Phi})$  and the standard deviation (SD) of parameters were calculated by

$$\bar{\Phi} = \frac{1}{B} \sum_{i=1}^{B} \Phi_i$$
$$SD(\Phi) = \sqrt{\frac{1}{B-1} \sum_{i=1}^{B} (\Phi_i - \bar{\Phi})^2}$$

where B is bootstrap resampling number.

The skewness (SK) and kurtosis (KT) are given by

$$SK(\Phi) = \frac{1/B \cdot \sum_{i=1}^{B} (\Phi_{i} - \bar{\Phi})^{3}}{\left(1/B \cdot \sum_{i=1}^{B} (\Phi_{i} - \bar{\Phi})^{2}\right)^{3/2}}$$
$$KT(\Phi) = \frac{1/B \cdot \sum_{i=1}^{B} (\Phi_{i} - \bar{\Phi})^{4}}{\left(1/B \cdot \sum_{i=1}^{B} (\Phi_{i} - \bar{\Phi})^{2}\right)^{2}}$$

When the histogram of a pharmacokinetic parameter s close to the normal distribution ( $SK \approx 0$  and  $KT \approx 3$ ), the normal distribution test was performed. When the histogram of a parameter is close to a log-normal distribution ( $SK \gg 0$  and  $KT \gg 3$ ), the logarithm of the parameter is used for the normal hypothesis test.

**Bailer's algorithm:** In Bailer's original paper,<sup>1)</sup> the mean and SE are discussed only for AUC. Therefore, the present discussion is the expansion of Bailer's

algorithm. The mean  $E(\Phi) = \overline{\Phi}$  and variance  $(V(\Phi))$  of MRT, CL and V<sub>ss</sub> in addition to those of AUC were calculated by the following equations which were derived according to the formula of four arithmetic operations of probability variable.

$$E(AUC) = \sum_{i=2}^{m} (E(C_i) + E(C_{i-1})) \cdot (t_i - t_{i-1})/2$$

$$V(AUC) = \sum_{i=2}^{m} (V(C_i) + V(C_{i-1})) \cdot ((t_i - t_{i-1})/2)^2$$

$$E(AUMC) = \sum_{i=2}^{m} (t_i \cdot E(C_i) + t_{i-1} \cdot E(C_{i-1})) \cdot (t_i - t_{i-1})/2$$

$$V(AUMC) = \sum_{i=2}^{m} (t_i^2 \cdot V(C_i) + t_{i-1}^2 \cdot V(C_{i-1})) \cdot (t_i - t_{i-1})/2)^2$$

$$E(MRT) = E(AUMC)/E(AUC)$$

$$V(MRT) = \frac{V(AUMC)}{E(AUC)^2} + \frac{V(AUC) \cdot E(AUMC)^2}{E(AUC)^4}$$

$$E(CL) = D/E(AUC)$$

$$V(CL) = D^2 \cdot V(AUC)/E(AUC)^4$$

$$E(V_{ss}) = E(CL) \cdot E(MRT)$$

$$V(V_{ss}) = E(MRT)^2 \cdot V(CL) + E(CL)^2 \cdot V(MRT)$$

where m is point number of time course (pseudoprofile),  $t_i$  and  $C_i$  are time and concentration, respectively, D is dose, and AUMC is area under moment curve. The time course is integrated by the trapezoidal formula without extrapolation. The standard deviation (SD) of each parameter is given as the root of the variance  $V(\Phi)$ .

 $E(C_i)$  and  $V(C_i)$  are the mean and variance of the concentrations at time  $t_i$ , respectively, and defined by

$$E(C_i) = \frac{1}{n_i} \sum_{j=1}^{n_i} C_{ij}$$
$$V(C_i) = \frac{1}{n_i - 1} \sum_{j=1}^{n_i} (C_{ij} - E(C_i))^2$$

where  $n_i$  is point number of concentrations at time  $t_i$ .

Now, we define the variance of mean  $(\Phi)$  by the following equations.

$$V(\overline{AUC}) = \sum_{i=2}^{m} (V(C_i)/n_i + V(C_{i-1})/n_{i-1})$$

$$\times ((t_i - t_{i-1})/2)^2$$

$$V(\overline{AUMC}) = \sum_{i=2}^{m} (t_i^2 \cdot V(C_i)/n_i + t_{i-1}^2 \cdot V(C_{i-1})/n_{i-1})$$

$$\times ((t_i - t_{i-1})/2)^2$$

$$V(\overline{MRT}) = \frac{V(\overline{AUMC})}{E(AUC)^2} + \frac{V(\overline{AUC}) \cdot E(AUMC)^2}{E(AUC)^4}$$

$$V(\overline{CL}) = D^2 \cdot V(\overline{AUC})/E(AUC)^4$$

$$V(\overline{V}_{ss}) = E(MRT)^2 \cdot V(\overline{CL}) + E(\overline{CL})^2 \cdot V(\overline{MRT})$$

where we note the relationship:  $E(\Phi) = E(\overline{\Phi})$ .

When  $n_i$  are the same (n), we find the following simple relationships:

$$V(\overline{AUC}) = \frac{1}{n} V(AUC)$$
$$V(\overline{AUMC}) = \frac{1}{n} V(AUMC)$$
$$V(\overline{CL}) = \frac{1}{n} V(CL)$$
$$V(\overline{V}_{ss}) = \frac{1}{n} V(V_{ss})$$

The standard error (SE) of each pharmacokinetic parameter is given as the root of the variance  $V(\bar{\Phi})$ .

The normal hypothesis test is carried out using the following equation.

$$Z_0 = \frac{|\bar{\Phi}_1 - \bar{\Phi}_2|}{\sqrt{SE_1^2 + SE_2^2}}$$

where  $\bar{\Phi}_1$  and  $\bar{\Phi}_2$  are means of a pharmacokinetic parameter, and  $SE_1$  and  $SE_2$  are standard errors in groups 1 and 2, respectively. If  $Z_0 > 1.96$  (confidence interval p<0.05), the difference is assumed to be significant between groups 1 and 2.

### **Results and Discussion**

Figure 1 shows the time courses of the plasma concentration of <sup>111</sup>In-Hsp70, Suc<sub>20</sub>-BSA, and Suc<sub>40</sub>-BSA after intravenous injection into mice, where each plot is expressed as the average ( $\pm$  standard deviation) of three mice.<sup>14,19</sup> The resampling number in the bootstrap calculation was considered for evaluating the optimum value. As a representative example, Fig. 2 shows the effect of the bootstrap resampling number (B) on the variance of parameters calculated by MOMENT(BS). Figures 2a and b show the averages and standard deviations of the mean(AUC) and mean(CL) versus B for three trial bootstrap calculations, whereas Figs. 2c and d show the averages and standard deviations of SD(AUC) and SD(CL) versus B. The means were almost constant and the standard deviations were close to zero with B of 200 or more, whereas the means and standard deviations exhibited marked fluctuations in B from 5- to 100-fold. Figure 3 shows statistical histograms of AUC (a, b, c) and CL (d, e, f) and the theoretical curves of the normal distribution of AUC with B values of 10, 200 and 1000, and Figs. 3d, e and f show the histograms and the theoretical curves of CL. The histograms at B (10) deviate from the theoretical curves of a normal distribution, whereas they are close to the theoretical curves at B values of 200 or 1000. Therefore, bootstrap resampling of 200 or more is enough to calculate the mean and SD, which is in good agreement with the conclusion by Efron et al.<sup>27,28)</sup> Considering these findings, B(1000) was



Fig. 1. Plasma concentration and liver accumulation of <sup>111</sup>In-Hsp70,  $Suc_{20}$ -BSA,  $Suc_{40}$ -BSA after intravenous injection into mice at several doses. These results are expressed mean ± standard deviation of three mice.



**Fig. 2.** Effect of bootstrap number on the estimated pharmacokinetic parameters. (a) and (b) represent the average of AUC and CL of <sup>111</sup>In-Hsp70 with standard deviation after intravenous injection at a dose of  $10 \,\mu\text{g/mouse}$ . (c) and (d) represent the average of SD with standard deviation, calculated at three trials by MOMENT(BS).

selected in the following calculations.

**Table 1** shows the means of AUC, MRT, CL and V<sub>ss</sub> and the standard deviations (SD) calculated by MOMENT(BS) after intravenous injection of <sup>111</sup>In-Hsp70, Suc<sub>20</sub>-BSA and Suc<sub>40</sub>-BSA into mice. The mean values and SD by bootstrap method were in good agreement with those calculated by Bailer's method. **Table 2** shows the skewness (SK) and kurtosis (KT) of the pharmacokinetic parameters of the three physiologically active proteins. All SK fluctuate around zero, whereas

all KT were about two to three. Based on the agreement between the bootstrap and Bailer's approaches and on the fact that SK were close to 0 and that KT were close to 3, all histograms of the parameters calculated by MOMENT(BS) were assumed to be normal (Gaussian) distributions, not log-normal distributions. These discussions are ignored in Bailer's approach. Thus, an attempt was made to compare the parameters between two groups under different conditions based on normal distribution, where it is noted that *SE* of parameters are



Fig. 3. (a), (b) and (c) represent the frequency histograms of AUC of Hsp70 after intravenous injection at a dose of  $10 \,\mu g/mouse$  into mice, whereas (d), (e) and (f) show the frequency of the CL at B numbers (10, 200 and 1000).

		AUC (µg/mL·min)				MRT (min)				CL (mL/min)				V <sub>ss</sub> (mL)			
Dose		Bootstrap		Bailer's		Bootstrap		Bailer's		Bootstrap		Bailer's		Bootstrap		Bailer's	
		Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD
Hsp70	$2 \mu g/mouse$	45.4	2.8	45.4	2.6	49.0	3.1	49.1	6.3	0.044	0.003	0.044	0.003	2.16	0.12	2.16	0.30
	$10 \mu\text{g/mouse}$ $100 \mu\text{g/mouse}$	195 2640	19 165	196 2640	18 157	68.0 53.4	6.8 3.1	67.6 53.4	9.8 6.4	0.052	0.005	0.051	0.005	3.53 2.03	0.61	3.44 2.02	0.59 0.27
Suc <sub>20</sub> -BSA	0.1 mg/kg	102	3	101	3	27.0	0.6	27.0	1.3	0.030	0.001	0.030	0.001	0.800	0.038	0.799	0.05
	1 mg/kg	673	8	673	8	22.7	0.3	22.7	0.7	0.046	0.001	0.045	0.001	1.01	0.02	1.01	0.03
	10 mg/kg	7850	261	7850	237	26.2	0.3	26.2	1.2	0.038	0.001	0.038	0.001	1.00	0.03	1.00	0.05
	20 mg/kg	16380	638	16400	686	28.2	0.9	28.1	2.4	0.037	0.001	0.037	0.002	1.03	0.03	1.03	0.10
Suc <sub>40</sub> -BSA	0.05 mg/kg	4.41	0.29	4.41	0.31	29.7	1.5	29.7	3.8	0.342	0.023	0.340	0.002	10.2	0.7	10.1	1.5
	0.1 mg/kg	9.56	0.62	9.57	0.62	24.7	1.4	24.7	3.1	0.315	0.021	0.314	0.002	7.76	0.60	7.73	1.1
	1 mg/kg	106	7	106	7	12.5	1.4	12.5	2.4	0.283	0.020	0.282	0.020	3.52	0.38	3.52	0.73
	10 mg/kg	3890	175	3900	173	18.8	1.0	18.8	1.9	0.077	0.004	0.077	0.003	1.45	0.06	1.45	0.16
	20 mg/kg	9760	364	9770	367	16.0	0.4	16.0	0.9	0.062	0.002	0.061	0.002	1.00	0.05	0.982	0.069

Table 1. Mean and Standard Deviation (SD) of AUC, MRT, CL and V<sub>ss</sub> calculated by MOMENT(BS)

given by  $SD/\sqrt{n}$ , respectively, in bootstrap and Bailer's approaches.

The difference of MRT of Hsp70 was insignificant between 2 and 100  $\mu$ g. The difference of SUC<sub>20</sub>-BSA was insignificant between 0.1 and 10 mg, between 0.1 and 20 mg, and between 10 and 20 mg. The difference of Suc<sub>40</sub>-BSA was insifnificant between 0.05 and 0.1 mg. The difference of MRT was significant among others.

The difference of CL of  $Suc_{20}$ -BSA was insignificant between 10 and 20 mg. The difference of  $Suc_{40}$ -BSA was

insignificant between 0.05 and 0.1 mg, and between 0.1 and 1 mg. The difference of CL was significant among others.

The difference of  $V_{ss}$  of Hsp-70 was insignificant between 2 and 100  $\mu$ g. The difference of Suc<sub>40</sub>-BSA was insignificant between 1 and 10 mg, between 1 and 20 mg, and between 1 and 20 mg. The difference of Vss was significant among others.

Generally, it was concluded that the pharmacokinetic parameters of Hsp70 and Suc<sub>20</sub>-BSA are almost

Dose -		AUG	C	MR	Г	CL		V <sub>ss</sub>		
		SK	KT	SK	KT	SK	KT	SK	KT	
Hsp70	$2 \mu g/mouse$	0.217	2.48	0.178	1.89	0.075	2.39	0.112	2.62	
	$10 \mu g/mouse$	0.155	2.14	0.265	2.28	0.190	2.15	0.200	2.01	
	$100 \mu g/mouse$	0.163	2.49	0.547	1.98	0.094	2.48	0.207	2.62	
	0.1 mg/kg	0.298	2.35	-0.099	2.30	-0.186	2.28	-0.355	2.13	
Suc DSA	1 mg/kg	-0.215	2.51	-0.255	1.99	0.686	2.67	0.180	2.74	
Suc <sub>20</sub> -BSA	10 mg/kg	0.301	2.00	0.164	2.23	-0.232	1.98	0.063	2.39	
	10 mg/kg 20 mg/kg - 0	-0.033	2.37	-0.327	1.83	0.179	2.42	-0.089	2.28	
	0.05 mg/kg	-0.335	2.72	-0.623	2.23	0.643	3.09	0.072	2.50	
Suc <sub>40</sub> -BSA	0.1 mg/kg	0.097	2.66	-0.107	2.17	0.221	2.62	0.401	2.70	
	1 mg/kg	0.244	2.48	-0.111	1.94	0.040	2.41	0.007	2.24	
	10 mg/kg	-0.253	2.33	0.007	2.20	0.441	2.49	0.133	2.70	
	20 mg/kg	0.356	2.12	0.069	2.26	-0.225	2.06	-0.326	1.91	

Table 2. Skewness (SK) and Kurtosis (KS) of AUC, MRT, CL and V<sub>ss</sub> calculated by MOMENT(BS)

independent of dose, whereas those of  $Suc_{40}$ -BSA are strictly dose-dependent.

The bootstrap method has been applied exclusively to estimating the confidence intervals of statistics ( $\Phi$ ).<sup>29-31)</sup> The interval analysis is equivalent to the test of statistical hypothesis. The test of hypothesis in addition to the histogram analysis by the bootstrap method is the first trial for the analysis of one-sampling data. The means and SD of the pharmacokinetic parameters estimated by the bootstrap method agreed with those obtained by Bailer's method, using MOMENT(BS). Consequently, SE by the bootstrap method were in good agreement with those by Bailer's method. The histograms of the parameters showed a normal distribution, which was close to the theoretical curve, and SK and KT were close to zero and three, respectively, for the present data using three physiologically active proteins. The proposed method is expected to be useful for assessing the histogram of a pharmacokinetic parameter. After assessment of the statistical distribution, it was shown that the hypothesis test can be applied to statistically comparing the disposition of two groups under different conditions, using one-point sampling data from animal experiments.

Acknowledgment: This work is partly supported by the 21st Century COE Program "Knowledge Information Infrastructure for Genome Science".

# References

- Bailer, A. J.: Testing for the equality of area under the curves when using destructive measurement techniques. *J. Pharmcokinetcis and Biopharmaceutics*, 16: 303–309 (1987).
- 2) Efron, B.: Bootstrap method: another look at the jacknife. *Annals Statistics*, **7**: 1–26 (1979).
- 3) Efron, B. and Tibshirani, R.: Statistical data analysis in the computer age. *Science*, **253**: 390–395 (1991).
- 4) Zellner, D., Frankewitsch, T. and Keller, F.: Synthesis

of pharmacokinetic parameters of vancomycin via bootstrap methods. Int. J. Clin. Pharmacol. Ther., 36: 554–560 (1998).

- 5) Yafune, A. and Ishiguro, M.: Bootstrap approach for constructing confidence intervals for population pharmacokinetic parameters. I: A use of bootstrap standard error. *Stat. Med.*, **18**: 581-591 (1999).
- Yafune, A. and Ishiguro, M.: Bootstrap approach for constructing confidence intervals for population pharmacokinetic parameters. II: a bootstrap modification of standard two-stage (STS) method for phase I trial. *Stat. Med.*, 18: 601-612 (1999).
- Tatami, S., Sarashina, A., Yamamura, N., Igarashi, T. and Tanigawara, Y.: Population of pharmacokinetics of an angiotensin II receptor antagonist, telmisartan, in healthy volunteers and hypertensive patients. *Drug. Metab. Pharmacokinet.*, 18: 203-211 (2003).
- Rajagopalan, P. and Gastonguay, M. R.: Population pharmacokinetics of ciprofloxacin in pediatric patients. *J. Clin. Pharmacol.*, 43: 698-710 (2003).
- Shao, J., Chow, S. C. and Wang, B.: The bootstrap procedure in individual bioequivalence. *Stat. Med.*, 19: 2741–2754 (2000).
- Kimanani, E. K.: Definition of individual bioequivalence: occation-to-occation versus mean switchability. *Stat. Med.*, 19: 2797–2810 (2000).
- 11) Kimanani, E. K.: Numerical methods for the evaluation of individual bioequivalence criteria. *Stat. Med.*, **19**: 2775–2795 (2000).
- Niselman, A. V., Garcia-Ben, M. and Rubio, M. C.: Robust methods in bioequivalence assay; preliminary results. *Eur. J. Drug. Metab. Pharmacokinet.*, 23: 148–152 (1998).
- Lange, K., Cantor, R., Horvath, S., Perola, M., Sabatti, C., Sinsheimer, J. and Sobel, E.: Mendel version 4.0: a complete package for the exact genetic analysis of discrete traits in pedigree and population data sets. Am. J. Hum. Genet., 69 (suppl.): A1886 (2001).
- 14) Takemoto, S., Nishikawa, M. and Takakura, Y.: Pharmacokinetic and tissue distribution mechanisms of mouse recombinant heat shock protein 70 in mice. *Pharm. Res.*, 22: 419-426 (2005).

- 15) Moroi, Y., Mayhew, M., Trcka, J., Hoe, M. H., Takechi, Y., Hartl, F. U., Rothman, J. E. and Houghton, A. N.: Induction of cellular immunity by immunization with novel hybrid peptides complexed to heat shock protein 70. *Proc. Natl. Acad. Sci. USA.*, 97: 3485–3490 (2000).
- Hartl, F. U.: Molecular chaperones in cellular protein folding. *Nature*, 381: 571–579 (1996).
- Udono, H. and Srivastava, P. K.: Comparison of tumorspecific immunogenicities of stress-induced proteins gp96, hsp90, and hsp70. J. Immunol., 152: 5398-5403 (1994).
- Srivastava, P. K.: Interaction of heat shock proteins with peptides and antigen presenting cells: chaperoning of the innate and adaptive immune responses. *Annu. Rev. Immunol.*, 20: 395-425 (2002).
- 19) Yamasaki, Y., Sumimoto, K., Nishikawa, M., Yamashita, F., Yamaoka, K., Hashida, M. and Takakura, Y.: Pharmacokinetic analysis of *in vivo* disposition of succinylated proteins targeted to liver nonparenchymal cells *via* scavenger receptors: importance of molecular size and negative charge density for *in vivo* recognition by receptors. *J. Pharmacol. Exp. Ther.*, 301: 467-477 (2002).
- 20) Yamasaki, Y., Hisazumi, J., Yamaoka, K. and Takakura, Y.: Efficient scavenger receptor-mediated hepatic targeting of proteins by introduction of negative charges on the proteins by aconitylation: the influence of charge density and size of proteins molecules. *Eur. J. Pharm. Sci.*, 18: 305-312 (2003).
- Srivastava, P. K.: Purification of heat shock proteinpeptide complexes for use in vaccination against cancers and intracellular pathogens. *Methods.*, 12: 165–171 (1997).
- 22) Shakushiro, K., Yamasaki, Y., Nishikawa, M. and Takakura, Y.: Efficient scavenger receptor-mediated up-

take and cross-presentation of negatively charged soluble antigens by dendritic cells. *Immunology*, **112**: 211–218 (2004).

- 23) Hnatowich, D. J., Layne, W. W. and Childs, R. L.: The preparation and labeling of DTPA-coupled albumin. *Int. J. Appl. Radiat Isot.*, **33**: 327-332 (1982).
- 24) Duncan, J. R. and Welch, M. J.: Intracellular metabolism of indium-111-DTPA-labeled receptor targeted proteins. J. Nucl. Med., 34: 1728–1738 (1993).
- 25) Arano, Y., Mukai, T., Uezono, T., Wakisaka, K., Motonari, H., Akizawa, H., Taoka, Y. and Yokoyama, A.: A biological method to evaluate bifunctional chelating agents to label antibodies with metallic radionuclides. J. Nucl. Med., 35: 890-898 (1994).
- Mager, H. and Goller, G.: Analysis of pseudo-profiles in organ pharmacokinetics and Toxicokinetics. *Stat. Med.*, 14: 1009–1024 (1995).
- Efron, B. and Tibshirani, R.: Bootstrap methods for standard erorrs, confidence intervals, and other measures of statistical accuracy. *Statistical. Science*, 1: 54–77 (1993).
- 28) Davison, A. C. and Hinkley, D. V.: Bootstrap methods and their application. *Cambridge University Press*, Cambridge (1997).
- 29) Pai, S. M., Fettner, S. H., Hajian, G., Cayen, M. N. and Batra, V. K.: Characterization of AUCs from sparsely sampled populations in toxicology studies. *Pharm. Res.*, 13: 1283-1290 (1996).
- 30) Bonate, P. L.: Coverage and precision of confidence intervals for area under the curve using parametric and non-parametric methods in a toxicokinetic experimental design. *Pharm. Res.*, 15: 405-410 (1998).
- Mager, H. and Goller, G.: Resampling methods in sparse sampling situations in preclinical pharmacokinetic studies. J. Pharm. Sci., 87: 372–378 (1998).