

Template for Reporting Results of Biomarker Testing of Specimens From Patients With Carcinoma of the Colon and Rectum

Template web posting date: December 2014

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For the Members of the Cancer Biomarker Reporting Workgroup, College of American Pathologists

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CAP Colon and Rectum Biomarker Template Revision History

Version Code

The definition of the version code can be found at www.cap.org/cancerprotocols.

Version: ColonBiomarkers 1.2.0.0

Summary of Changes

The following changes have been made since the October 2013 release.

RESULTS

Microsatellite Instability

Format for percentage reporting was changed and reference was added for National Cancer Institute (NCI) markers.

Loci Testing

An option for "Not performed" was added.

KRAS Mutational Analysis

Reference to "wild type KRAS allele" was removed. "Mutation not stated" was revised to "mutation, not otherwise specified." Reporting option for "GIn61Leu (CAA>CCA)" was added.

NRAS Mutational Analysis BRAF Expression (by immunohistochemistry) Sections were added.

BRAF Mutational Analysis

Reference to "wild type BRAF allele" was removed. Formatting for reporting "BRAF V600E (c.1799T>A) mutation" and "Other BRAF mutation" was revised.

PIK3CA Mutational Analysis

Reference to "wild type PIK3CA allele" was removed.

PTEN Mutational Analysis

Reference to "wild type PTEN allele" was removed.

METHODS

KRAS Mutational Analysis Options for specifying applicable codons were removed.

NRAS Mutational Analysis

Section was added.

PTEN Expression and Mutational Analysis

Options added to specify IHC antibody and ISH probe used in testing.

Biomarker Reporting Template

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Completion of the template is the responsibility of the laboratory performing the biomarker testing and/or providing the interpretation. When both testing and interpretation are performed elsewhere (eg, a reference laboratory), synoptic reporting of the results by the laboratory submitting the tissue for testing is also encouraged to ensure that all information is included in the patient's medical record and thus readily available to the treating clinical team.

COLON AND RECTUM

Select a single response unless otherwise indicated.

Note: Use of this template is optional.

+ RESULTS

- + Immunohistochemistry (IHC) Testing for Mismatch Repair (MMR) Proteins (select all that apply) (Note A)
- + ____ MLH1
 - + ____ Intact nuclear expression
 - + ____ Loss of nuclear expression
 - + ____ Cannot be determined (explain): _____
- + ____ MSH2
 - + ____ Intact nuclear expression
 - + ____ Loss of nuclear expression
 - + ____ Cannot be determined (explain): _____
- + ____ MSH6
 - + ____ Intact nuclear expression
 - + ____ Loss of nuclear expression
 - + ____ Cannot be determined (explain): _____
- + ____ PMS2
 - + ____ Intact nuclear expression
 - + ____ Loss of nuclear expression
 - + ___ Cannot be determined (explain): ___
- + ____ Background nonneoplastic tissue/internal control with intact nuclear expression

+ IHC Interpretation

- + ____ No loss of nuclear expression of MMR proteins: low probability of microsatellite instability-high (MSI-H)#
- + _____Loss of nuclear expression of MLH1 and PMS2: testing for methylation of the *MLH1* promoter and/or mutation of *BRAF* is indicated (the presence of a *BRAF* V600E mutation and/or *MLH1* methylation suggests that the tumor is sporadic and germline evaluation is probably not indicated; absence of both *MLH1* methylation and of *BRAF* V600E mutation suggests the possibility of Lynch syndrome, and sequencing and/or large deletion/duplication testing of germline *MLH1* may be indicated)#

- + ____ Loss of nuclear expression of MSH2 and MSH6: high probability of Lynch syndrome (sequencing and/or large deletion/duplication testing of germline *MSH2* may be indicated, and, if negative, sequencing and/or large deletion/duplication testing of germline *MSH6* may be indicated)#
- + ____ Loss of nuclear expression of MSH6 only: high probability of Lynch syndrome (sequencing and/or large deletion/duplication testing of germline *MSH6* may be indicated)[#]
- + ____ Loss of nuclear expression of PMS2 only: high probability of Lynch syndrome (sequencing and/or large deletion/duplication testing of germline PMS2 may be indicated)#

[#] There are exceptions to the above IHC interpretations. These results should not be considered in isolation, and clinical correlation with genetic counseling is recommended to assess the need for germline testing.

+ Microsatellite Instability (MSI) (Note A)

- + ____ MSI stable (MSS)
- + ____ MSI low (MSI-L)
 - + ____ 1% 29% of the National Cancer Institute (NCI) or mononucleotide markers exhibit instability
 - + ____ 1 of the NCI or mononucleotide markers exhibit instability
 - + ____ Other (specify): _____
- + ____ MSI high (MSI-H)
 - + ___ ≥30% of the NCI or mononucleotide markers exhibit instability
 - + ____ 2 or more of the NCI or mononucleotide markers exhibit instability
 - + ____ Other (specify): _____
- + ____ MSI indeterminate

+ Loci Testing

- + ____ Mononucleotide panel
 - + BAT-25
 - + ____ Stable
 - + ____ Unstable
 - + ___ Cannot be determined (explain): _____
 - + ____ Not performed
 - + BAT-26
 - + ____ Stable
 - + ____ Unstable
 - + ___ Cannot be determined (explain): _____
 - + ____ Not performed
 - + NR-21
 - + ____ Stable
 - + ____ Unstable
 - + ____ Cannot be determined (explain): _____
 - + ____ Not performed
 - + NR-24
 - + ____ Stable
 - + ____ Unstable
 - + ____ Cannot be determined (explain): _____
 - + ____ Not performed
 - + Mono-27
 - + ____ Stable
 - + ____ Unstable
 - + ____ Cannot be determined (explain): _____
 - + ____ Not performed

- + ____ NCI panel
 - + BAT-25
 - + ____ Stable
 - + ____ Unstable
 - + ___ Cannot be determined (explain): _____
 - + ____ Not performed
 - + BAT -26
 - + ____Stable
 - + ____ Unstable
 - + ___ Cannot be determined (explain): _____
 - + ____ Not performed
 - + D2S123
 - + ____ Stable
 - + ____ Unstable
 - + ___ Cannot be determined (explain): _____
 - + ____ Not performed
 - + D5S346
 - + ____ Stable
 - + ____ Unstable
 - + ___ Cannot be determined (explain): _____
 - + ____ Not performed
 - + D17S250
 - + ____ Stable
 - + ____ Unstable
 - + ___ Cannot be determined (explain): _____
 - + ____ Not performed
- + ____ Other (specify): ______
 - + ____ Stable
 - + ____ Unstable
 - + ___ Cannot be determined (explain): _____

+ MLH1 Promoter Methylation Analysis (Note B)

- + ____ MLH1 promoter hypermethylation present
- + ____ MLH1 promoter hypermethylation absent
- + ___ Cannot be determined (explain): _____

+ KRAS Mutational Analysis (Note C)

- + ____ No mutation detected
- + ____ Mutation identified (select all that apply)
 - + Codon 12
 - + ____ Gly12Asp (GGT>GAT)
 - + ____ Gly12Val (GGT>GTT)
 - + ____ Gly12Cys (GGT>TGT)
 - + ____ Gly12Ser (GGT>AGT)
 - + ____ Gly12Ala (GGT>GCT)
 - + ____ Gly12 Arg (GGT>CGT)
 - + ____ Codon 12 mutation, not otherwise specified
 - + ___ Other codon 12 mutation (specify): _____
 - + Codon 13
 - + ____ Gly13Asp (GGC>GAC)
 - + ____ Gly13Arg (GGC>CGC)
 - + ____ Gly13Cys (GGC>TGC)

- + ____ Gly13Ala (GGC>GCC)
- + ____ Gly13Val (GGC>GTC)
- + ____ Codon 13 mutation, not otherwise specified
- + ____ Other codon 13 mutation (specify): _____
- + Codon 61
- + ____ Gln61Leu (CAA>CTA)
- + ____ GIn61His (CAA>CAC)
- + ____ Codon 61 mutation, not otherwise specified
- + ____ Other codon 61 mutation (specify): ______
- + Codon 146
- + ____ Ala146Thr (G436A) (GCA>ACA)
- + ____ Codon 146 mutation, not otherwise specified
- + ___ Other codon 146 mutation (specify): _____
- + ____ Other codon (specify): ______
- + ___ Cannot be determined (explain): _____

+ NRAS Mutational Analysis (Note C)

- + ____ No mutation detected
- + ____ Mutation identified (select all that apply)
 - + Codon 12
 - + ____ Gly12Asp (GGT>GAT)
 - + ____ Gly12Val (GGT>GTT)
 - + ____ Gly12Cys (GGT>TGT)
 - + ____ Gly12Ser (GGT>AGT)
 - + ____ Gly12Ala (GGT>GCT)
 - + ____ Gly12Arg (GGT>CGT)
 - + ____ Codon 12 mutation, not otherwise specified
 - + ____ Other codon 12 mutation (specify): _____
 - + Codon 13
 - + ____ Specific codon 13 mutation (specify): ____
 - + ____ Codon 13 mutation, not otherwise specified
 - + Codon 61
 - + ____ GIn61Lys (CAA>AAA)
 - + ____ Gln61Arg (CAA>CGA)
 - + ____ Codon 61 mutation, not otherwise specified
 - + ___ Other codon 61 mutation (specify): _____
 - + ____ Other codon (specify): ______
- + ___ Cannot be determined (explain): _____

+ BRAF Expression (by immunohistochemistry) (Note B)

- + ____ Positive cytoplasmic expression
- + ____ Negative for cytoplasmic expression
- + ___ Cannot be determined (explain): _____

+ BRAF Mutational Analysis (Note B)

- + ____ No mutations detected
- + ____ BRAF V600E (c.1799T>A) mutation
- + ____ Other BRAF mutation (specify): _
- + ___ Cannot be determined (explain): _____

+ PIK3CA Mutational Analysis (Note D)

- + ____ No mutations detected
- + ____ Exon 9 mutation present (specify): _____
- + ____ Exon 20 mutation present (specify): _____
- + ___ Cannot be determined (explain):

+ PTEN Expression (by immunohistochemistry) (Note E)

- + ____ Positive cytoplasmic and/or nuclear expression
- + ____ Negative for cytoplasmic and nuclear expression
- + ___ Cannot be determined (explain): _____

+ PTEN Mutational Analysis

- + ____ No mutation detected
- + ____ Exon 1-9 mutation present (specify): _____
- + ____ Cannot be determined (explain): _____

+ Multiparameter Gene Expression/Protein Expression Assay

- + Specify type: _____
- + Results:
- + ____ Low risk
- + ____ Moderate risk
- + ____ High risk
- + Recurrence score: _____

+ METHODS

+ Dissection Method(s) (select all that apply) (Note F)

- + ____ Laser capture microdissection
- + Specify test name#: _____
- + <u>Manual under microscopic observation</u> + Specify test name[#]:
 - + specify test name^{π}: ___
- + ____ Manual without microscopic observation
 - + Specify test name#: _____
- + ____ Cored from block
 - + Specify test name[#]: _____
- + ____ Whole tissue section (no tumor enrichment procedure employed)
 - + Specify test name#: _____

[#] If more than 1 dissection method used, please specify which test was associated with each selected dissection method.

+ Microsatellite Instability (MSI)

- + Number of MSI markers tested (specify): _____
- + <u>Cellularity</u>
- + Percent tumor cells present in specimen: _____%

+ Whole Genome or Exome Sequencing

- + ____ Whole genome sequencing (specify): ______
- + ____ Whole exome sequencing (specify): _____

+ MLH1 Promoter Methylation

- + Testing Method
- + ____ Methylation-specific real-time polymerase chain reaction (PCR)
- + ____ Other (specify): ______

+ KRAS Mutational Analysis

- + <u>Codons Assessed</u> (select all that apply)
- + ____ 12
- + ____ 13
- + ____ 61
- + ____ 146
- + <u>Testing Method(s)</u> (select all that apply)
- + ____ Direct Sanger sequencing
- + ____ Pyrosequencing
- + ____ High-resolution melting analysis
- + ____ PCR, allele-specific hybridization
- + ____ Real-time PCR
- + ___ Other (specify): _____

Please specify in Comments section if different testing methods are used for different codons.

+ NRAS Mutational Analysis

- + Codons Assessed (select all that apply)
- + ____ 12
- + ____ 13
- + ____ 61

+ <u>Testing Method(s)</u> (select all that apply)

- + ____ Direct Sanger sequencing
- + ____ Pyrosequencing
- + ____ High-resolution melting analysis
- + ____ PCR, allele-specific hybridization
- + ____ Real-time PCR
- + ____ Other (specify): _____

Please specify in Comments section if different testing methods are used for different codons.

+ BRAF Mutational Analysis

- + Mutations Assessed (select all that apply)
- + ____ V600E
- + ____ Other BRAF V600 mutation (specify): _____
- + ____ Other (specify): _____

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- + Testing Method (select all that apply)
- + ____ Direct Sanger sequencing
- + ____ PCR, allele-specific hybridization
- + ____ Pyrosequencing
- + ____ Real-time PCR
- + ____ Immunohistochemistry for V600E gene product
- + ____ Other (specify): ______

+ PIK3CA Mutational Analysis

- + Testing Method
- + ____ Direct Sanger sequencing
- + ____ Other (specify): ______

+ PTEN Expression and Mutational Analysis

- + <u>Testing Method</u> (select all that apply)
- + ____ Immunohistochemistry (specify antibody): _____
- + ___ In situ hybridization (specify probe): _____
- + ____ Direct Sanger sequencing
- + ____ Duplication/deletion testing (MLPA)
- + ____ Other (specify): _____

+ COMMENT(S)

Note: Fixative type, time to fixation (cold ischemia time), and time of fixation should be reported if applicable in this template or in the original pathology report.

Gene names should follow recommendations of The Human Genome Organisation (HUGO) Nomenclature Committee (www.genenames.org; accessed February 10, 2015).

All reported gene sequence variations should be identified following the recommendations of the Human Genome Variation Society (www.hgvs.org/mutnomen/; accessed February 10, 2015).

Explanatory Notes

A. Mismatch Repair Testing: Microsatellite instability and Immunohistochemistry

Detection of defective mismatch repair in colorectal carcinomas is important for detection of Lynch syndrome (hereditary nonpolyposis colorectal cancer syndrome [HNPCC]), which accounts for approximately 2% to 3% of all colorectal carcinomas and has clinical implications for treatment of the affected patient and family members. Microsatellite instability (MSI) testing can be used to cost-effectively screen colorectal cancer patients for possible Lynch syndrome. Patients with a microsatellite instability-high (MSI-H) phenotype that indicates mismatch repair deficiency in their cancer may have a germline mutation in one of several DNA mismatch repair (MMR) genes (eg, *MLH1, MSH2, MSH6*, or *PMS2*) or an altered *EPCAM (TACSTD1)* gene. After appropriate genetic counseling, patients may want to consider testing to identify the causative heritable abnormality. An MSI-H phenotype is more frequently observed in sporadic colorectal cancer (about 15% of cases) due to somatic abnormalities, usually hypermethylation of the *MLH1* gene promoter. The specificity of MSI testing can be increased by using it primarily on at-risk populations, such as colorectal cancer patients younger than 50 years, or patients with a strong family history of Lynch-associated tumors (eg, colorectal, endometrial, gastric, or upper urinary tract urothelial carcinoma),¹ but with sacrifice of sensitivity, since a sizeable minority of cases lacks these clinical characteristics.

MSI testing of tumor DNA is generally performed with at least 5 microsatellite markers, generally mononucleotide or dinucleotide repeat markers. In 1998, a National Institutes of Health consensus panel proposed that laboratories use a 5-marker panel consisting of 3 dinucleotide and 2 mononucleotide repeats for MSI testing.² Recent data suggests that dinucleotide repeats may have lower sensitivity and specificity for identifying tumors with an MSI-H phenotype. As a consequence, there has been a move towards including more mononucleotides and fewer dinucleotides in MSI testing panels. Many laboratories now use a commercially available kit for MSI testing that utilizes 5 mononucleotide markers.

MSI testing is frequently done in conjunction with immunohistochemical (IHC) testing for DNA MMR protein expression (ie, MLH1, MSH2, MSH6, and PMS expression). If DNA MMR IHC has not been performed, this testing should be recommended for any case that shows an MSI-H phenotype, because this information will help identify the gene that is most likely to have a germline mutation (eg, a patient whose tumor shows loss of MSH2 and MSH6 expression, but retention of MLH1 and PMS2 expression, is likely to have an *MSH2* germline mutation). If the results of DNA MMR IHC and MSI testing are discordant (eg, MSI-H phenotype with normal IHC or abnormal IHC with MSS phenotype), then the laboratory should make sure that the same sample was used for MSI and IHC testing and that there was no sample mix-up. However, MSI-H may not occur in colorectal cancers of patients with germline *MSH6* mutation. Intact expression of all 4 proteins indicates that MMR enzymes tested are intact but does not entirely exclude Lynch syndrome, as approximately 5% of families may have a missense mutation (especially in *MLH1*) that can lead to a nonfunctional protein with retained antigenicity. Defects in lesser-known MMR enzymes may also lead to a similar result, but this situation is rare.

Any positive reaction in the nuclei of tumor cells is considered as intact expression (normal), and it is common for intact staining to be somewhat patchy. An interpretation of expression loss in tumor cells should be made only if a positive reaction is seen in internal control cells, such as the nuclei of stromal, inflammatory, or nonneoplastic epithelial cells. Loss of expression of MLH1 may be due to Lynch syndrome or methylation of the *MLH1* promoter region (as occurs in sporadic MSI colorectal carcinoma). Genetic testing is ultimately required for this distinction, although a specific *BRAF* gene mutation (V600E) is present in many sporadic cases, but not familial cancers. Loss of MSH2 expression strongly suggests Lynch syndrome. PMS2 loss is often associated with loss of MLH1 and is only independently meaningful if MLH1 is intact. MSH6 is similarly related to MSH2. One should also keep in mind that nucleolar staining or complete loss of MSH6 staining has been described in colorectal cancer

cases with prior radiation or chemotherapy,^{3,4} and a significant reduction of MSH6 staining has been described in a small percentage of colorectal carcinomas with somatic mutations of the coding region microsatellites of the *MSH*6 gene in MLH1/PMS2-deficient carcinomas.⁵

B. MLH1 Promoter Hypermethylation Analysis and BRAF Mutational Analysis

Defective mismatch repair in sporadic colorectal cancer is most often due to inactivation of the *MLH1* gene promoter by hypermethylation (epigenetic silencing). The V600E mutation of the *BRAF* gene may be present in up to 70% of tumors with hypermethylation of the *MLH1* promoter. In colorectal cancer, this mutation has been associated with a limited clinical response to epidermal growth factor receptor (*EGFR*) targeted therapies (cetuximab or panitumumab). Analysis for somatic mutations in the V600E hot spot in *BRAF* may also be indicated for tumors that show MSI-H, as this mutation has been found in sporadic MSI-H tumors, but not in Lynch-associated cancers with *MLH1* or *MSH2* mutations.⁶ *BRAF* V600E mutations have been described in probands with monoallelic *PMS2* mutations.⁷ Direct testing of *MLH1* promoter hypermethylation and/or the use of *BRAF* V600E mutational analysis prior to germline genetic testing in patients with MSI-H tumors for whom further testing is not indicated.⁸

C. RAS Mutational Analysis

The presence of a *KRAS* mutation has been shown to be associated with lack of clinical response to therapies targeted at *EGFR*, such as cetuximab⁹ and panitumumab.¹⁰ While clinical guidelines for *KRAS* mutational analysis are evolving, current provisional recommendations from the American Society of Clinical Oncology are that all patients with stage IV colorectal carcinoma who are candidates for anti-EGFR antibody therapy should have their tumor tested for *KRAS* mutations.¹¹ Anti-EGFR antibody therapy is not recommended for patients whose tumors show mutations in *KRAS* codon 12, 13, or 61, but data on codon 146 are currently insufficient. A recent study has shown that *NRAS* mutation, like *KRAS* mutation, has influence on response to anti-EGFR therapy.¹² Although more studies are needed, these findings may lead to broad *KRAS* and *NRAS* panels to include codons 12, 13, 61, and 146 of both genes.

D. PIK3CA Mutational Analysis

PIK3CA mutations activate the *PI3K-PTEN-AKT* pathway that is downstream from both the *EGFR* and the *RAS-RAF-MAPK* pathways. *PIK3CA* mutation and subsequent activation of the *AKT* pathway has been shown to play an important role in colorectal carcinogenesis and have been associated with *KRAS* mutation¹³ and microsatellite instability.¹⁴ *PIK3CA* mutation has further been associated with poor survival in resectable stage I to III colon cancer, with the adverse effect of *PIK3CA* mutation potentially limited to patients with *KRAS* wild-type tumors.¹⁵ *PIK3CA* mutations have been associated with resistance to anti-EGFR therapy in several studies,^{16,17} but not in others.¹⁸ The reasons for the discrepancy are not clear. Mutations of exons 1, 9, and 20 of the *PIK3CA* gene represent >95% of known mutations.

A European consortium recently suggested that only *PIK3CA* exon 20 mutations are associated with a lack of cetuximab activity in *KRAS* wild-type tumors and with a shorter median progression-free survival and overall survival.¹⁷ By contrast, exon 9 *PIK3CA* mutations are associated with *KRAS* mutations and do not have an independent effect on cetuximab efficacy.¹⁷ More studies are needed to establish the prognostic and predictive roles of *PIK3CA* exon-9 and exon-20 mutations.

E. PTEN Mutational Analysis

The role of *PTEN* loss in colorectal cancer prognosis and therapy is unclear. It has been suggested that loss of *PTEN* expression, as determined by immunohistochemistry, is associated with lack of benefit from cetuximab in metastatic colorectal cancer.¹⁹⁻²² Loss of *PTEN* has been found to co-occur with *KRAS*, *BRAF*, and *PIK3CA* mutations.^{19,22} The recorded frequency of loss of *PTEN* expression varies from 19% to 36%, with some studies reporting an effect on response rate and survival, whereas others found an effect only on progression-free or overall survival. Moreover, data on the loss of *PTEN* expression are not

concordant in primary and metastatic tissues.²¹ There is currently no standardized method for PTEN expression analysis by immunohistochemistry.

F. Dissection Method

Please denote the manner in which the tissue was dissected and specify the biomarker test only if different dissection methods are used for different tests.

- 1. Laser capture microdissection (LCM): Use of a laser-equipped microscope to isolate and retrieve specific cells of interest from a histopathologic region of interest.
- 2. Manual under microscopic observation: hematoxylin and eosin (H&E) slide is examined under a light microscope and marked by a pathologist for subsequent tumor dissection and retrieval.
- 3. Manual without microscopic observation: H&E slide is examined without a microscope and marked by a pathologist for subsequent tumor dissection and retrieval.
- 4. Cored from block: Area of interest is cored from a paraffin-embedded tissue block.
- 5. Whole tissue section: No tumor enrichment procedure employed for tissue retrieval.

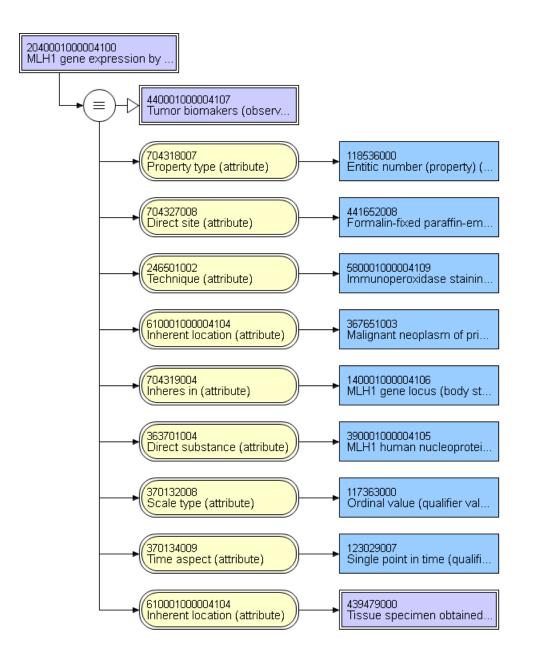
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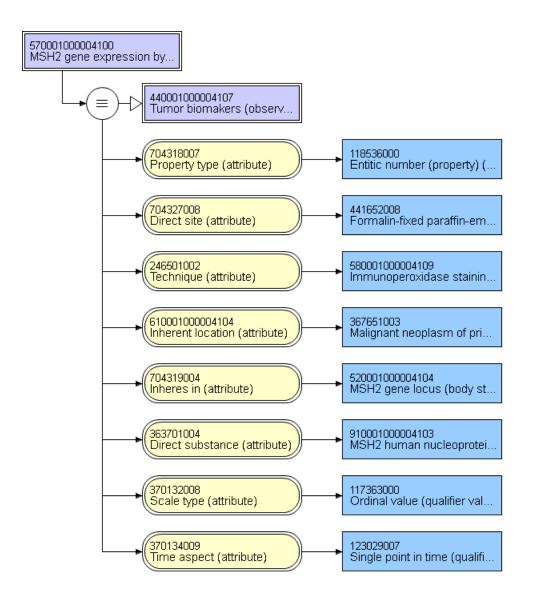
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This ends the CAP cancer protocols. Following pages represents suggested LOINC and SNOMED CT value sets for use with protocols.

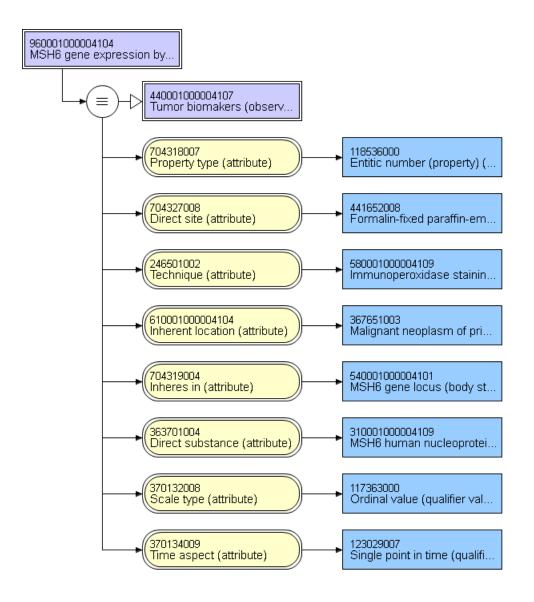
MLH1 Analysis	81691-8 - DNA mismatch repair pr Immune stain	81691-8 - DNA mismatch repair protein Mlh1 [Presence] in Cancer by Immune stain	
	Intact nuclear expression	2090001000004107 Intact genetic nucleoprotein expression (finding)	
	Loss of nuclear expression	1810001000004106 Loss of genetic nucleoprotein expression (finding)	
	Indeterminate	82334004 Indeterminate (qualifier value)	



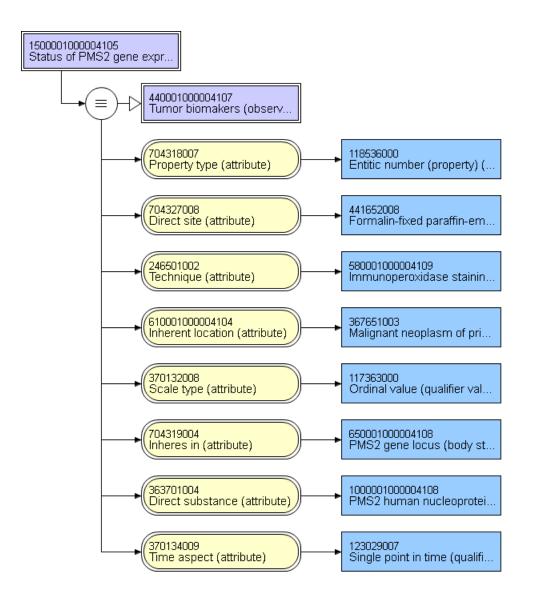
MSH2 Analysis	81692-6 - DNA mismatch repair pr Immune stain	81692-6 - DNA mismatch repair protein Msh2 [Presence] in Cancer by Immune stain	
	Intact nuclear expression	2090001000004107 Intact genetic nucleoprotein expression (finding)	
	Loss of nuclear expression	1810001000004106 Loss of genetic nucleoprotein expression (finding)	
	Indeterminate	82334004 Indeterminate (qualifier value)	



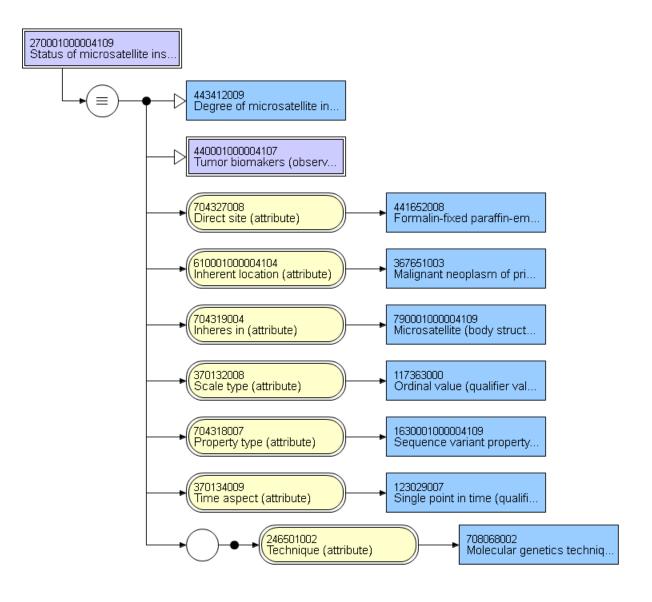
MSH6 Analysis	81693-4 - DNA mismatch repair protein Msh6 [Presence] in Cancer by Immune stain	
	Intact nuclear expression	2090001000004107 Intact genetic nucleoprotein expression (finding)
	Loss of nuclear expression	1810001000004106 Loss of genetic nucleoprotein expression (finding)
	Indeterminate	82334004 Indeterminate (qualifier value)



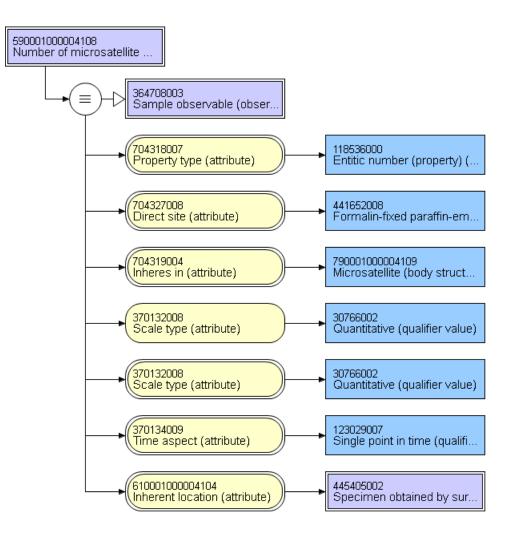
PMS2 Analysis	81694-2 - Mismatch repair endon Immune stain	81694-2 - Mismatch repair endonuclease PMS2 [Presence] in Cancer by Immune stain	
	Intact nuclear expression	2090001000004107 Intact genetic nucleoprotein expression (finding)	
	Loss of nuclear expression	1810001000004106 Loss of genetic nucleoprotein expression (finding)	
	Indeterminate	82334004 Indeterminate (qualifier value)	



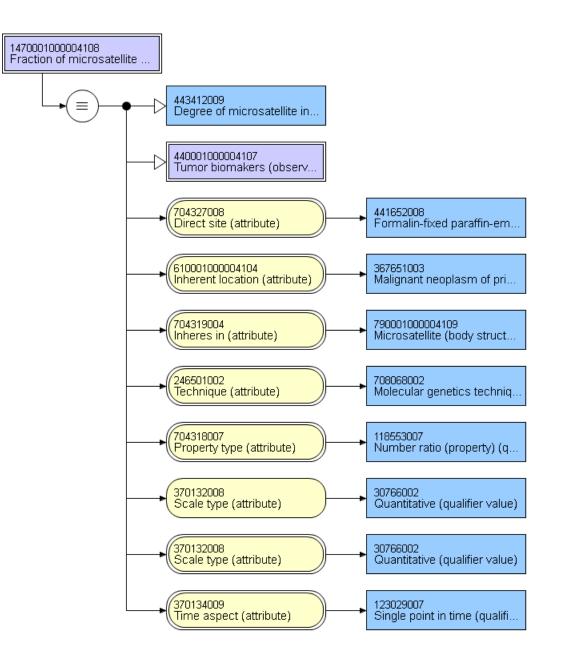
Microsatellite instability	81695-9 - Microsatellite instability [Interpretation] in Cancer Qualitative	
	Stable (MSS)	58158008 Stable (qualifier value)
	Low (MSI-L)	62482003 Low (qualifier value)
	High (MSI-H)	75540009 High (qualifier value)
	Indeterminate	82334004 Indeterminate (qualifier value)



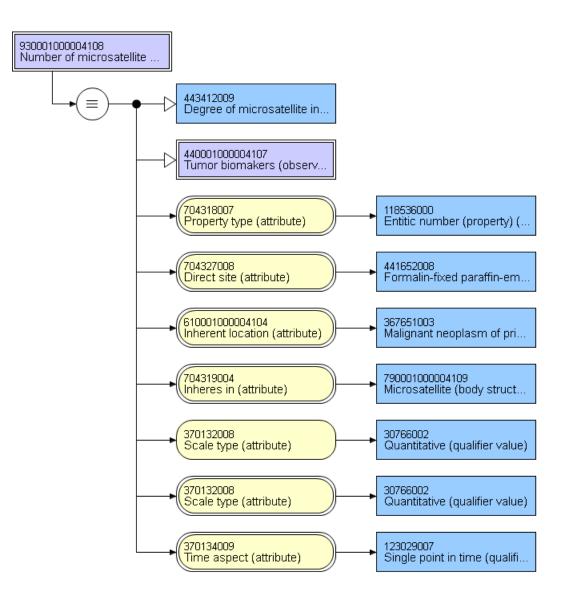
Number of microsatellite markers	81696-7 - Microsatellite instability markers assessed [#] in	
assessed	Cancer	



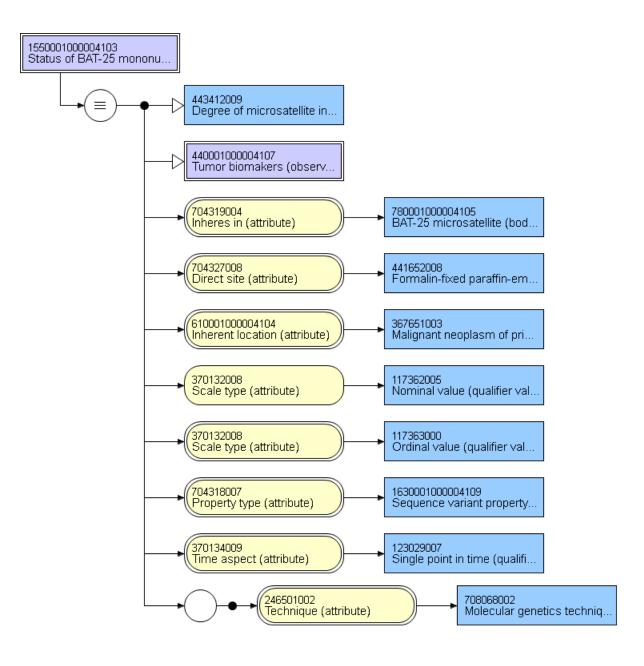
Percent microsatellite markers exhibiting instability	81708-0 - Microsatellite markers exhibiting instability/Microsatellite instability markers assessed in Cancer	
	%	



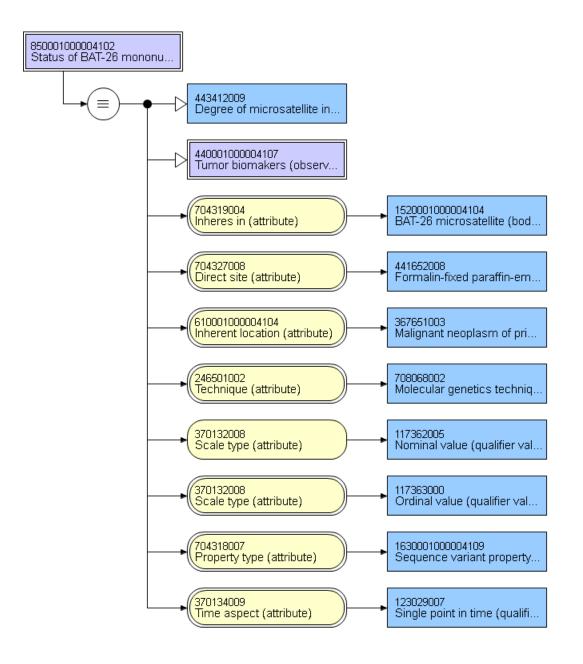
Number of microsatellite markers exhibiting instability	81709-8 - Microsatellite markers exhibiting instability [#] in Cancer	



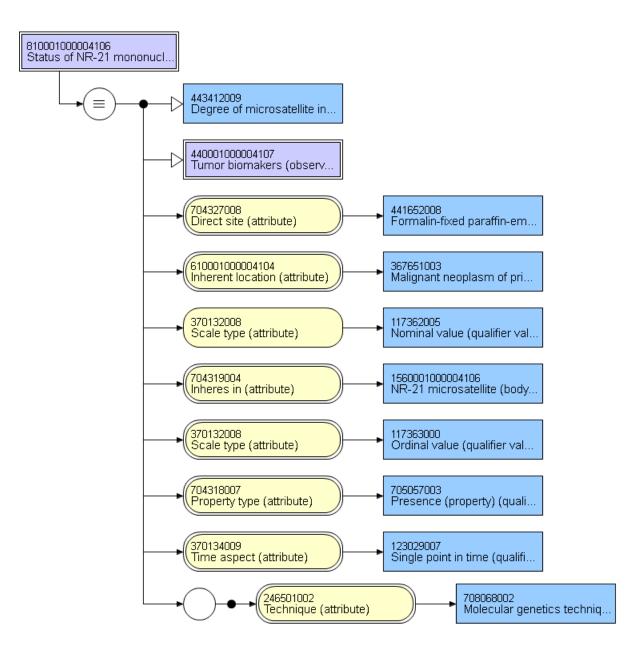
BAT-25 Loci stability	81697-5 - Microsatellite instability marker BAT25 in Cancer Qualitative	
	Stable	58158008 Stable (qualifier
		value)
	Unstable	263922001 Unstable status
		(qualifier value)
	Undetermined	373068000 Undetermined
	(explain):	(qualifier value)



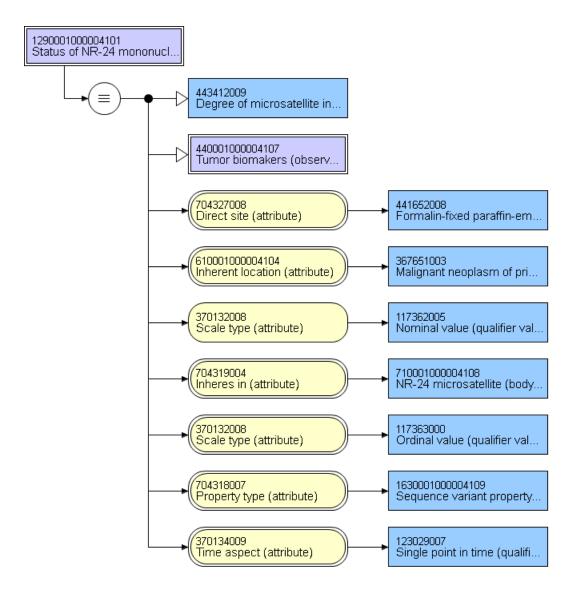
BAT-26 Loci stability	81698-3 -	
	Microsatellite instability marker BAT26 in Cancer Qualitative	
	Stable	58158008 Stable (qualifier value)
	Unstable	263922001 Unstable status (qualifier
		value)
	Undetermined	373068000 Undetermined (qualifier
	(explain):	value)



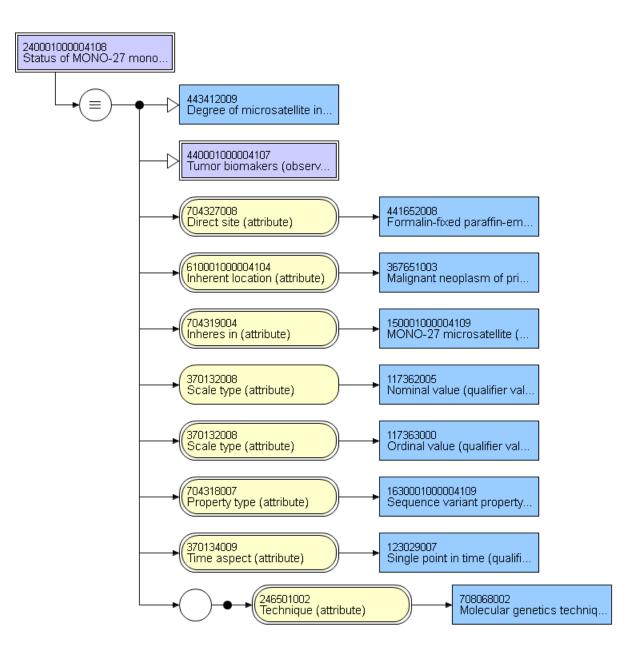
NR-21 Loci	81699-1 -		
stability	Microsatellite instability marker NR21 in Cancer Qualitative		
	Stable	Stable 58158008 Stable (qualifier value)	
	Unstable	263922001 Unstable status	
		(qualifier value)	
	Undetermined (explain):	373068000 Undetermined (qualifier	
		value)	



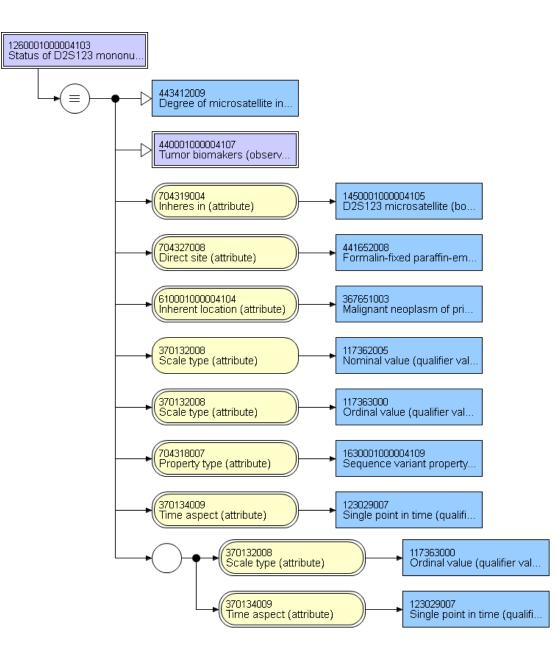
NR-24 Loci stability	81700-7 - Microsatellite instability marker NR24 in Cancer Qualitative	
	Stable	58158008 Stable
		(qualifier value)
	Unstable	263922001 Unstable
		status (qualifier value)
	Undetermined (explain):	373068000
		Undetermined (qualifier
		value)



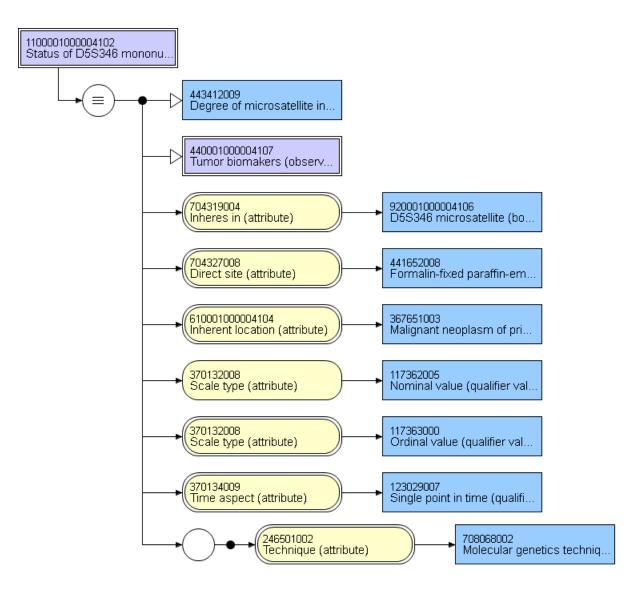
Mono-27	81701-5 -				
Loci stability	Microsatellite instability marker MONO27 in Cancer Qualitative				
	Stable 58158008 Stable (qualifier value)				
	Unstable	263922001 Unstable status (qualifier			
		value)			
	Undetermined (explain):	373068000 Undetermined (qualifier			
		value)			



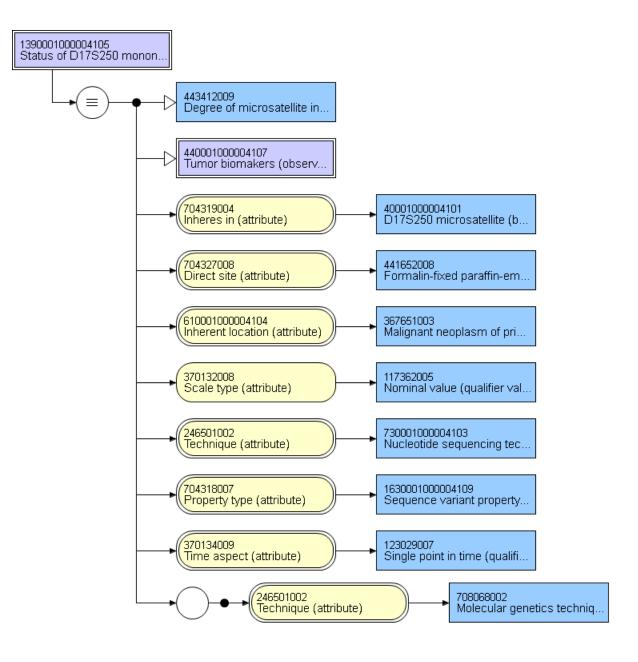
D2S123 Loci stability	81702-3 - Microsatellite instability marker D2S123 in Cancer Qualitative		
	Stable	58158008 Stable (qualifier value)	
	Unstable	263922001 Unstable status (qualifier value)	
	Undetermined	373068000 Undetermined (qualifier value)	
	(explain):		



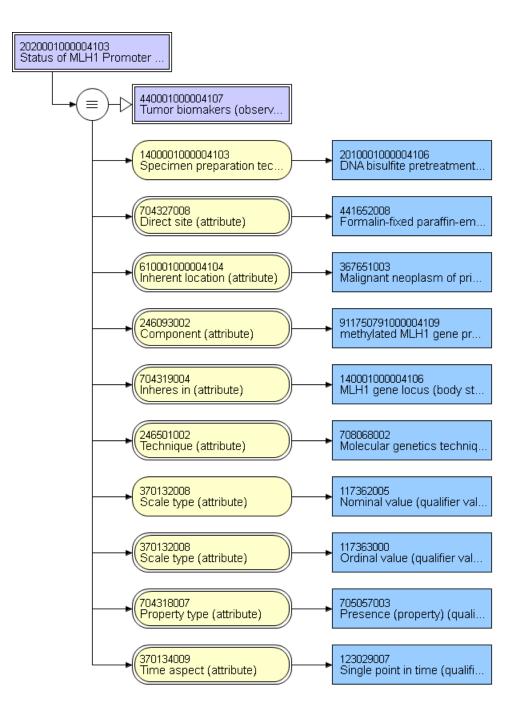
D5S346 Loci stability	8	81703-1 - Microsatellite instability marker D5S346 in Cancer Qualitative			
		Stable 58158008 Stable (qualifier value)			
		Unstable	263922001 Unstable status (qualifier		
			value)		
		Undetermined (explain):	373068000 Undetermined (qualifier		
		value)			



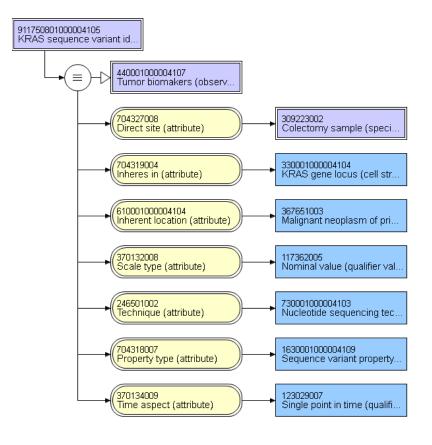
D17S250 Loci stability	81704-9 - Microsatellite instability marker D17S250 in Cancer Qualitative			
	Stable 58158008 Stable (qualifier value)			
	Unstable	263922001 Unstable status (qualifier value)		
	Undetermined (explain):	373068000 Undetermined (qualifier value)		



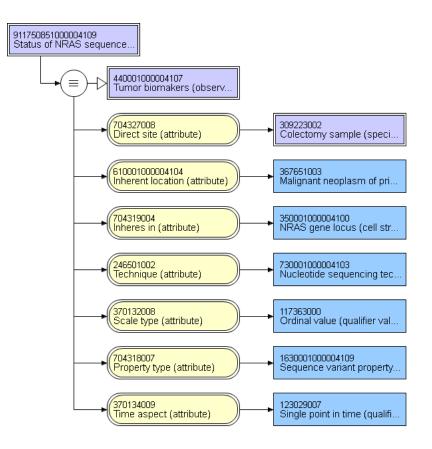
MLH1 Promotor Methylation Analysis	58416-9 - MLH1 gene methylation [Presence] in Tissue by Molecular genetics method			
	MLH1 promoter hypermethylation 52101004 Present (qualifier			
	present	value)		
	MLH1 promoter hypermethylation	2667000 Absent (qualifier		
	absent	value)		
	Undetermined (explain):	373068000 Undetermined		
	(qualifier value)			



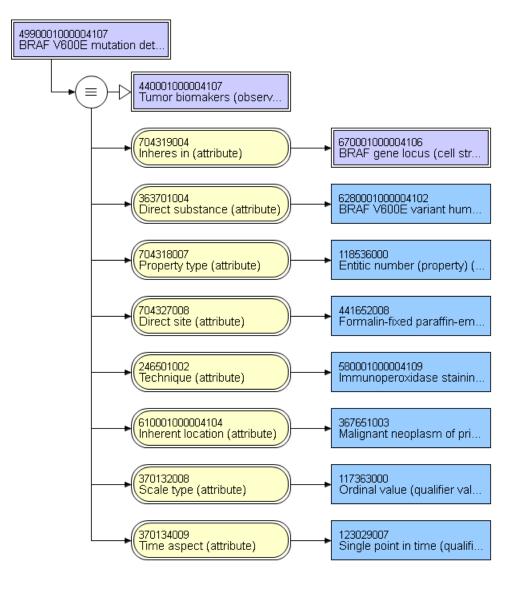
KRAS Mutation		
Identified		
	No sequence variant identified	NM_004985 c.(=)
	Gly12Asp (GGT>GAT)	NM_004985.4(KRAS):c.35G>A (p.Gly12Asp)
	Gly12Val (GGT>GTT)	
	Gly12Cys (GGT>TGT)	
	Gly12Ser (GGT>AGT)	
	Gly12Ala (GGT>GCT)	
	Gly12Arg (GGT>CGT)	
	Other Codon 12 mutation (specifiy):	
	Gly13Asp (GGC>GAC)	
	Gly13Arg (GGC>CGC)	
	Gly13Cys (GGC>TGC)	
	Gly13Ala (GGC>GCC)	
	Gly13Val (GGC>GTC)	
	Other Codon 13 mutation (specifiy):	
	GIn61Leu (CAA>CTA)	
	Gln16His (CAA>CAC)	
	Other Codon 16 mutation (specifiy):	
	Ala146Thr (G435A) (GCA>ACA)	
	Other Codon 146 mutation	
	(specifiy):	
	Other Codon mutation (specify):	



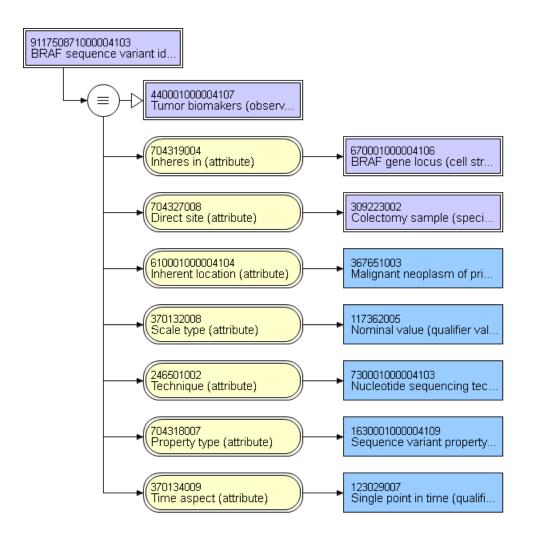
NRAS Mutation Identified		
	Gly12Asp (GGT>GAT)	
	Gly12Val (GGT>GTT)	
	Gly12Cys (GGT>TGT)	
	Gly12Ser (GGT>AGT)	
	Gly12Ala (GGT>GCT)	
	Gly12Arg (GGT>CGT)	
	Other Codon 12 mutation (specifiy):	
	Other Codon 13 mutation (specifiy):	
	Gin61Lys (CAA>AAA)	
	GIn61Arg (CAA>CGA)	
	Other Codon 61 mutation (specifiy):	
	Other Codon mutation (specify):	



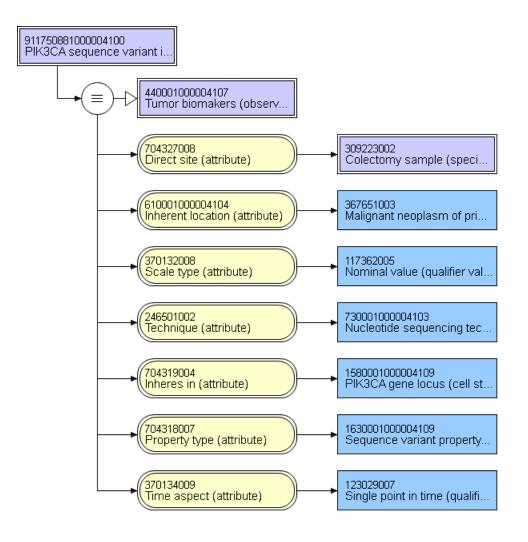
BRAF Expression by IHC	81705-6 - Serine-threonine protein kinase B-raf [Presence] in Cancer by Immune stain		
	Positive cytoplasmic expression	318120301000004107 Positive genetic protein expression in cytoplasm of cell (finding)	
	Negative cytoplasmic expression	307815691000004106 Loss of genetic protein expression in cytoplasm of cell (finding)	
	Undetermined (explain) :	373068000 Undetermined (qualifier value)	



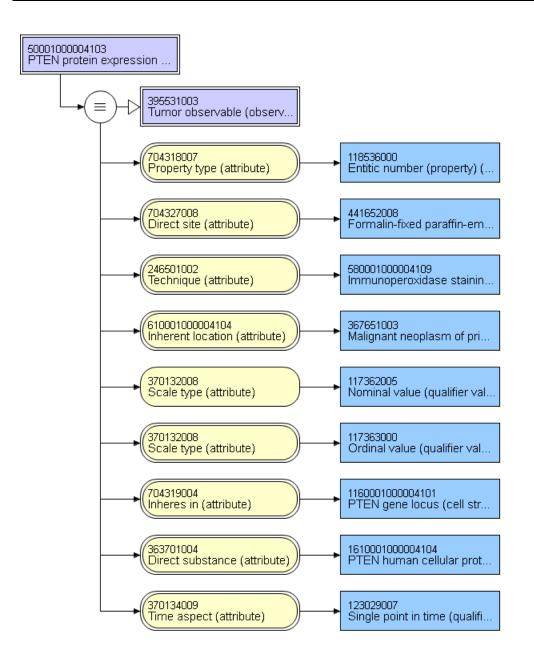
BRAF Mutational identified		
	BRAF V600E mutation	843509901000004102 BRAF V600E mutation identified in excised malignant neoplasm (finding)
	Other BRAF mutation (specify):	



PIK3CA Mutation Identified		
	Exon 9 mutation (specify):	
	Exon 20 mutation (specify):	



PTEN Expression by IHC	81706-4 - Phosphatase and tensin homolog (PTEN) protein [Presence] in Cancer by Immune stain		
	Positive for cytoplasmic and/or nuclear expression	826864521000004106 Positive gene expression in cell (finding)	
	Negative for cytoplasmic and/or nuclear expression	62582271000004100 Negative gene expression in cell (finding)	
	Undetermined (explain) :	373068000 Undetermined (qualifier value)	



PTEN Mutation Identified		
	Exon 1-9 mutation (specify):	

